The Patent Office
Concept House
Cardiff Road
Newport
South Wales
NP10 8QQ

I, the undersigned, being an officer duly authorised in accordance with Section 74(1) and (4) of the Deregulation & Contracting Out Act 1994, to sign and issue certificates on behalf of the Comptroller-General, hereby certify that annexed hereto is a true copy of the documents as originally filed in connection with the patent application identified therein.

I also certify that the attached copy of the request for grant of a Patent (Form 1/77) bears an amendment, effected by this office, following a request by the applicant and agreed to by the Comptroller-General.

In accordance with the Patents (Companies Re-registration) Rules 1982, if a company named in this certificate and any accompanying documents has re-registered under the Companies Act 1980 with the same name as that with which it was registered immediately before re-registration save for the substitution as, or inclusion as, the last part of the name of the words "public limited company" or their equivalents in Welsh, references to the name of the company in this certificate and any accompanying documents shall be treated as references to the name with which it is so re-registered.

In accordance with the rules, the words "public limited company" may be replaced by p.l.c., plc, P.L.C. or PLC.

Re-registration under the Companies Act does not constitute a new legal entity but merely subjects the company to certain additional company law rules.

Signed

Dated 26 April 2004

PRIORITY DOCUMENT
SUBMITTED OR TRANSMITTED IN COMPLIANCE WITH RULE 17.1(a) OR (b)

Patents Form 1/77

nts Act 1977
le 16)

THE PATENT OFFICE
A
5 APR 2003
NEWPORT

1/77

Request for grant of a patent

(See the notes on the back of this form. You can also get an explanatory leaflet from the Patent Office to help you fill in this form)

5 APR 2003

The Patent Office

Cardiff Road
Newport
South Wales
NP10 8QQ

1. Your reference ~~PC/JM/P12531GB~~ ORF/BP62156S1

07APR03 E790264 1 D00230
P01/7700 0.00-0307969.6

2. Patent application number *(The Patent Office will fill in this part)* 0307969.6

3. Full name, address and postcode of the or of each applicant *(underline all surnames)*

THE UNIVERSITY COURT OF THE UNIVERSITY OF GLASGOW
Gilbert Scott Building
University Avenue
Glasgow G12 8QQ UNITED KINGDOM

Patents ADP number *(if you know it)* 7738460003

If the applicant is a corporate body, give the country/state of its incorporation

4. Title of the invention "Living Tissue Model & Uses Thereof"

5. Name of your agent *(if you have one)* ~~CRUIKSHANK & FAIRWEATHER~~ MEWBURN ELLIS LLP

"Address for service" in the United Kingdom to which all correspondence should be sent *(including the postcode)* ~~19 Royal Exchange Square Glasgow G1 3AE UNITED KINGDOM~~ York House 23 Kingsway London WC2B 6HP

Patents ADP number *(if you know it)* ~~547002~~ ✓ 109006

PFS1/77 5/4/04 WSA

6. If you are declaring priority from one or more earlier patent applications, give the country and the date of filing of the or of each of these earlier applications and *(if you know it)* the or each application number

Country	Priority application number *(if you know it)*	Date of filing *(day / month / year)*

7. If this application is divided or otherwise derived from an earlier UK application, give the number and the filing date of the earlier application

Number of earlier application	Date of filing *(day / month / year)*

8. Is a statement of inventorship and of right to grant of a patent required in support of this request? *(Answer 'Yes' if:*

a) any applicant named in part 3 is not an inventor, or
b) there is an inventor who is not named as an applicant, or
c) any named applicant is a corporate body.
See note (d))

Patents Form 1/77

9. Enter the number of sheets for any of the following items you are filing with this form. Do not count copies of the same document

Continuation sheets of this form	
Description	26
Claim(s)	-
Abstract	-
Drawing(s)	10 + 10

10. If you are also filing any of the following, state how many against each item.

Priority documents	
Translations of priority documents	
Statement of inventorship and right to grant of a patent *(Patents Form 7/77)*	
Request for preliminary examination and search *(Patents Form 9/77)*	
Request for substantive examination *(Patents Form 10/77)*	
Any other documents *(please specify)*	

11. I/We request the grant of a patent on the basis of this application.

Signature Cruikshank & Fairweather Date 4/4/03

CRUIKSHANK & FAIRWEATHER 4TH APRIL 2003

12. Name and daytime telephone number of person to contact in the United Kingdom

DR. P. CHAPMAN - 0141-221-5767

Warning

After an application for a patent has been filed, the Comptroller of the Patent Office will consider whether publication or communication of the invention should be prohibited or restricted under Section 22 of the Patents Act 1977. You will be informed if it is necessary to prohibit or restrict your invention in this way. Furthermore, if you live in the United Kingdom, Section 23 of the Patents Act 1977 stops you from applying for a patent abroad without first getting written permission from the Patent Office unless an application has been filed at least 6 weeks beforehand in the United Kingdom for a patent for the same invention and either no direction prohibiting publication or communication has been given, or any such direction has been revoked.

Notes

a) *If you need help to fill in this form or you have any questions, please contact the Patent Office on 08459 500505.*

b) *Write your answers in capital letters using black ink or you may type them.*

c) *If there is not enough space for all the relevant details on any part of this form, please continue on a separate sheet of paper and write "see continuation sheet" in the relevant part(s). Any continuation sheet should be attached to this form.*

d) *If you have answered 'Yes' Patents Form 7/77 will need to be filed.*

e) *Once you have filled in the form you must remember to sign and date it.*

f) *For details of the fee and ways to pay please contact the Patent Office.*

LIVING TISSUE MODEL & USES THEREOF

Introduction

The present invention relates to a method of testing agents for their effect on a murine living tissue model, wherein the model represents the progression from normal to benign or malignant tumour tissue. The model is particularly useful in evaluating any therapeutic or oncogenic properties of a test agent. The invention also relates to murine living tissue models for use in methods of the invention.

Background to the Invention

Conventional animal tests employed to evaluate new therapeutic anti-cancer agents or identify suspect carcinogens are expensive, time consuming, require skilled animal-trained staff and utilise large numbers of animals. To date *in vitro* alternatives have relied on the use of conventional cell culture systems which are limited in that they do not allow the three-dimensional interactions that occur between the tumour cells and with their surrounding stromal tissue. This is a serious disadvantage as such interactions are well documented as having a significant influence on the growth and invasion profiles of tumours.

EP0358506 relates to a three-dimensional cell and tissue culture system, based upon a synthetic mesh support on which cells are grown, which may be used in cytotoxicity testing of drugs. There is not however any suggestion of how to test oncogenic properties of a drug and/or potential usefulness of drugs as anti-cancer agents.

It is an object of the present invention to obviate and/or mitigate at least one of the aforementioned disadvantages. It is a further object of the present invention to provide an *in vitro* method suitable to allow evaluation of test compounds for oncogenic or anti-cancer properties that can, in part, replace the need to test in live animals.

The present invention is based on developing a model using a combination of murine epithelial (tumour) cells and stromal cells within a three-dimensional collagen gel that mimics a connective tissue matrix. Thus, models incorporate the influence of activated stromal cells on the growth and invasion characteristics of specific tumour cell lines following treatment with novel drugs or exposure to carcinogens in a similar fashion to epithelial tumours *in vivo*. The approach described herein comprises a new reproducible method, capable of incorporating computer analysis, of cell growth and invasion. This may be used to highlight the flexibility

of this *in vitro* tumour progression model in the assessment of the role, as an example, of retinoids, well established therapeutics, and additional genotoxic carcinogens or intrinsic mutations.

Thus, the present invention provides an *in vitro* method for observing an effect a test agent has on a mouse living tumour model, comprising the steps of:

a) providing at least one three-dimensional mouse living tissue model, wherein said model is intended to model normal, benign tumour or malignant tumour tissue;

b) contacting the test agent with said model(s); and

c) observing the effect the test agent has on said model(s).

Mouse cells are used in this model because of the importance of the mouse in studies of carcinogenesis. The model can be constructed of cells from specific genetically modified mice, thereby making use of this important, expanding animal resource, while minimising the numbers of animals needed. The models can utilise a variety of normal, benign and malignant mouse epithelial cell lines combined with mouse embryonic or neonatal fibroblasts.

The present *in vitro* model is intended in part to replace or reduce existing tests carried out on live mice via a pre-screening service. It is likely that some testing on living mice will still need to be conducted to

validate results as a companion test, but the intention is that this will be reduced.

The test agent may be any agent including chemical agents, pharmaceuticals, peptides, proteins, gene medicines, introduced genes or the like. The models described herein may be used to test for an agent's anti-cancer properties or alternatively for any carcinogenic properties of the test agent. As will be described later in more detail, the present inventors have developed "normal", "benign tumour" and "malignant tumour" models, which are particularly useful in testing a test agent's properties.

The mouse living tissue model may be a modified form of established systems which have been used for constructing human dermal equivalents. However, the living tumour tissue model is developed from and comprises mouse cells rather than human cells, as this is a more equivalent replacement to the living mouse carcinogenesis models currently used. The present invention is generally therefore directed to the development of mouse epithelial models responsible for all carcinomas. Typically this may include models of skin, mammary, intestinal or lung epithelial tissue (tumours).

A preferred model for use in the present invention comprises a disc, plug or the like of a collagen gel,

which may be formed, for example, formed from a solution of collagen into which fibroblasts are mixed. Once it has set, the fibroblasts contract the gel into a connective tissue like disc. Thereafter the contracted collagen gel or sponge is inoculated/seeded with epithelial (tumour) cells which adhere to the surface of the collagen or invade into the gel, forming tumour-like clusters or an epithelium characteristic of the tissue of origin. In another embodiment, fibroblasts and tumour cells are incorporated into the gel from the start, before it sets and contracts. The epithelial (tumour) cells and optionally the fibroblasts are derived from the appropriate tissue on which the model is to be based. That is, for example, if the model is a skin tumour model, the epithelial cells and optionally the fibroblasts are obtained from a source of skin tissue. Further incubation, either submerged or semi-submerged at body temperature for up to three weeks, allows the epithelial cells to grow and establish structures representative of the tumour of origin or normal tissue *in vivo*.

In an embodiment of the present invention, the present inventors have developed mouse skin tumour models in which newborn/embryonic mouse skin fibroblasts have been used to produce a contracted collagen gel. This collagen gel has then been utilised to produce three

distinct models - "normal", "benign" and "malignant" epidermal cell tumours.

The mouse epithelial cell line BalbMK may be used to produce, for example, a normal "control" *in vitro* model. The mouse skin epidermal papilloma cell line SP-1, which carries a mutant c-ras^{Ha} gene, may be used for example, to produce a benign or papilloma model when incorporated into the model. The T52 Hufos cell line is a variant of SP-1, formed from SP-1 cells which had been transfected with human fos and may be used to produce, for example, a model that is representative of an invasive malignant stage of tumour development. It will be appreciated however, that other suitable cell lines which develop "normal", "benign" or "malignant" models may be utilised. These may be originally developed from, for example, experimentally induced tumours in epidermis of mice carrying specific genetic alterations, for example, activated oncogenes or deleted tumour suppressor genes.

It is to be understood that a "normal" model is intended to be equivalent to the tissue architecture from which the cells are taken. Thus, using skin keratinocytes on a collagen gel impregnated with fibroblasts, leads to a model with a histopathology of a stratified epithelium on a normal dermis. Initially keratinocytes attach to the matrix and once raised to the air/liquid interface they form a basement membrane and

begin to differentiate forming a normal epidermis (illustrated in Figure 2). A "benign" model is intended to be equivalent to tissue in which a benign tumour, for example a papilloma, has developed. For example, the epithelial cells may clump together and grow together at the surface of the collagen, forming papilloma-like structures (see Figure 3). Finally, a "malignant" tumour model is a model in which the epithelial cells display an invasive nature and infiltrate the collagen gel, such that the epithelial cells do not just remain exposed at the surface of the collagen gel (see Figures 1 and 4).

Thus in accordance with the method of the present invention it is preferable that the test agent is tested on two of the models described herein, eg. the normal and malignant, or all three types of model ie. "normal", "benign" and "malignant". In this manner it is possible to determine a test agent's effect on different stages of tumour progression.

The test agent may be added to said model to be tested by any suitable means. For example, the test agent may be added drop-wise onto the epithelial surface of the model and allowed to diffuse into or otherwise enter the model, or it can be added to the nutrient medium and diffuse through the collagen gel to the epithelial/tumour cells. The model is also suitable for testing the effects of physical agents such as ionising

radiation, UV-light or heat alone or in combination with chemical agents (for example, in photodynamic therapy). Multiple models may be set up in, for example, multiwell tissue culture plates, to allow testing of many agents and/or different concentrations under different conditions.

Observing the effect the test agent has on said models may include a variety of methods. For example any changes in cell area or morphology (such as a cell entering apoptosis) may be observed directly on the intact model utilising for example suitable fluorescent cell staining. This can be by pre-labelling of tumour cells with vital dyes or genetically introduced fluorescent markers (for example green fluorescent proteins) for serial analysis of the living model or by fixation and post-labelling with fluorescent substances such as propidium iodide or fluorescently labelled antibodies. Alternatively, models may be processed by normal histological methods. Moreover, this may be carried out in an automated/robotic or semi-automated manner, using computer systems and software to image the cells at various time points and detect any change in, for example, cell density, location and/or morphology. Confocal laser scanning microscopy in particular permits three-dimensional analysis of intact models. Thus it is possible to apply directly to the intact, three-

dimensional tumour model, quantitative analysis of cell behaviour which are normally only possible for cells in conventional two-dimensional culture. By this means quantitative, serial analysis of cell proliferation, apoptosis, necrosis, migration and matrix invasion, among others, are obtained in a three-dimensional tumour cell model which bridges the gap between conventional two-dimensional cell cultures and live animal models.

Also, by appropriate control of viewing/photographing the model, such as by viewing/photographing several fields at random and thereafter randomly selecting a subset of these, it is possible to minimise any bias which may be introduced by a person analysing the data. It is also possible to observe if the test agent induces or inhibits cellular production of proteins, using suitable techniques known in the art, for example, using immunohistochemisty, immunofluorescence, PCR, microarrays, immunoblotting and zymography.

The present invention will now be further described by way of non-limiting examples only and with reference to the Figures which show:

Figure 1 shows a schematic representation of a model and how it may be utilised according to the present invention.

Figure 2: BalbMK "control" mouse immortalised epidermal keratinocyte model: Propidium iodide stained whole mounts (x 100 magnification) of (A) 2 day submerged culture and (B) 9 day submerged culture and H & E stained paraffin sections (x 100 magnification) of (C) 2 day submerged culture and (D) 10 day culture which has been raised to the air interface for 4 days.

Figure 3: SP-1 "papilloma" model: Propidium iodide stained whole mounts (x 100 magnification) of (A) 2 day submerged culture (B) 8 day submerged culture (C) 9 day culture which has been raised to the air interface for 1 day and H & E stained paraffin sections of (D) a 2 day submerged culture (x 100 magnification) and (E) a 10 day culture which has been raised to the air interface for 4 days, (F) reconstructed three-dimensional confocal image of a model as in (E) showing SP-1 cells covering the gel surface and piling up into papilloma-like structures.

Figure 4: T52 Hufos "invasive tumour" model: Propidium iodide stained whole mounts (x 100 magnification) of (A) 4 day submerged culture (B) 6 day submerged culture (C) 10 day culture that has been raised to the air interface for 4 days and H & E paraffin stained sections (x 400 magnification) of (D) 4 day submerged culture (E) 6 day submerged culture and (F) 10 day culture which has been raised to the air interface for 4 days.

Figure 5: The effect of 10^{-7}M retinoic acid on the area of cell cover in the SP-1 papilloma model over a 12 day period, assessed by imaging propidium iodide stained whole mounts.

Figure 6: The effect of retinoic acid concentration on the area of cell cover in the SP-1 papilloma model at day 8 and day 10 of culture.

Figure 7: The effect of 10^{-7}M retinoic acid on the area of cell cover in the T52 Hufos invasive tumour model over a 10 day culture period.

Figure 8: Detection of apoptotic cells. Condensed and fragmented cell nuclei, characteristic of cell death by apoptosis, can be detected on (A) an H & E stained paraffin section of a T52 Hufos model (x 400 magnification) and (B) a propidium iodide whole mount of an SP-1 model (x 200 magnification).

Figure 9 shows vital dye stained whole mounts (x 100 magnification) (A, C and E) and H & E stained paraffin sections (x 400 magnification) (B, D and F) of submerged cultures of CMT93/69 mouse rectum carcinoma (A & B), CMT64/61 mouse lung carcinoma (C & D) and TA3 Hauschka mouse mammary carcinoma (E and F).

Examples Section

Example 1: Preparation of the living tissue model

In summary the *in vitro* models were developed using contracted collagen gels which supported a layer of mouse tumour cells or corresponding "normal" epithelial cells. The collagen gel was comprised of type I collagen, isolated from rat tail tendon, contracted using primary mouse fibroblasts, which were isolated from the dermis of newborn (or embryonic) mice. The collagen gels contracted to a size approximately 1.5cm in diameter and were seeded with a single cell suspension of mouse tumour or normal epithelial cells. The models were initially maintained as submerged cultures, which allowed the cells to adhere to the collagen gel and grow. The models may thereafter be raised to the air-liquid interface (semi-submerged culture) to promote cell differentiation and formation of tissue. Benign tumour cells were observed to grow on the surface of the lattice and aggregate to form piles of cells equivalent to wart-like skin papillomas whereas, malignant tumour cells were observed to grow both on top of and into the support matrix mimicking invasive carcinomas. Test agents can be added to the model system at different stages of tumour development. The models are directly imaged as whole mounts by fluorescent labelling of cells either with

vital dyes or genetic markers or after fixation and staining.

In more detail, the collagen gel, described bellow, which supports the layer of epithelial cells (BalbMK (Weissman and Aaronson 1985), SP-1 (Strickland *et al.* 1988) or T52Hufos (Greenhalgh and Yuspa 1988) in the prototype models), was contracted using dermal fibroblasts isolated from newborn mouse skin. The epidermis and dermis were separated from each other following an overnight digestion with trypsin at 4°C. The dermal tissue was washed in sterile PBS, dissected into very small pieces, suspended in 3-5 ml of MEM culture media and the slurry seeded into a 75cm^2 plastic culture flask. This tissue was cultured without disturbance until the tissue pieces had adhered to the plastic flask. Fresh MEM (Minimum Essential Medium) supplemented with 10% foetal calf serum, 1% L-glutamine, 100U/ml penicillin and 100µg/ml streptomycin was added to the flask at this time. Fibroblast cell outgrowth from the dermal tissue explants was observed after several days. The excess tissue was removed and the fibroblast cells cultured to confluence. Collagen gels were prepared by mixing type I collagen solution, with ten-fold concentrated MEM and foetal calf serum containing mouse fibroblast cells in a ratio of 4:1:1. The

fibroblast density and collagen I concentration can be varied.

The collagen gels contracted from 3.5cm diameter to 1.5cm diameter using the optimised fibroblast seeding density of 0.3 x 10^6 fibroblasts per ml of collagen gel cast. Collagen gels were placed into 24 well culture dishes and seeded with epithelial cells. Cell seeding densities were optimised so that the epithelial cells produced measurable areas of cell cover. The seeding density was optimised to 0.5 x 10^6 cells/collagen gel. This seeding density ensures that there are sufficient cells present to form a good area of cell cover on the collagen lattice at the time of seeding. This is important, as the cells require cell-cell contact for further growth and differentiation on the collagen lattice support.

Models were routinely cultured for four days as submerged cultures. The models were then raised to the air interface by placing them onto porous stainless steel mesh or sintered glass supports with media reaching to the base of the epithelial cell layer. This allowed the model to continue to receive the nutrients from the culture medium as well as the support and the nutrients generated from the collagen lattice.

Three different mouse skin epithelial cell lines were used to create prototype *in vitro* models with

different properties. The BalbMK cells, which are slow growing mouse immortalised keratinocytes, grow in medium with a low concentration of calcium (0.05mM) supplemented with 5 ng/ml epidermal growth factor (EGF)(Weissman and Aaronson 1985). BalbMK cells, although immortalised, represent a normalised epithelial cell line and are used in the *in vitro* model system to provide a "normal", control model. The mouse skin epidermal papilloma cell line SP-1 carries a mutant *c-ras*^{Ha} gene. SP-1 cells grow in a low calcium environment and have papilloma-like qualities and are used to provide a benign tumour model (Strickland *et al.* 1988). The T52 Hufos cell line is a variant of SP-1 cells, which has been transfected with human fos, and grows in the same media as SP-1 cells supplemented with G418 and are used to provide a malignant tumour model (Greenhalgh and Yuspa 1988)

The models may easily be adopted to provide other models comprising alternative mouse epithelial cell lines of, for example, skin, mammary, intestinal or lung origin.

Example 2: Processing of models and data interpretation

Cell models were harvested at appropriate time intervals and fixed overnight at room temperature in a solution of buffered formalin. A small piece of the model was removed and embedded in paraffin wax. Sections

were cut and mounted onto glass slides and stained with haematoxylin and eosin (H & E stain). The remainder of the model was washed in PBS (phosphate buffered saline), permeabilised with Triton X-100 and stained with propidium iodide (PI). The washed models were stored thereafter in the dark at 4°C and maintained their fluorescence for several weeks. Whole mounts of the PI stained models were analysed using fluorescent microscopy through coverslips applied directly to the surface of the tissue.

Propidium iodide staining of the whole mounts allows changes in the cell nuclei to be observed. Condensation and fragmentation of the cell nucleus, indicative of cell death by apoptosis can be clearly identified (Figure 8). Haematoxylin/eosin staining identified changes in cell morphology, cell spreading, differentiation and cell death. Alternatively, tumour cells were pre-labelled with the fluorescent dye "DiI" before incorporation into the model. Labelled tumour cells could then be imaged directly in the living model (Figure 9).

The model is also suitable for applying immunohistological methods of detection to look for specific proteins, which may be altered by a specific test reagent.

Four random fields of view were selected for each model and photographed at x100 magnification. The areas

of cell cover were measured using the computer graphics package Adobe Photoshop. Images were downloaded as picture files directly into this programme. The PI stains all the cell nuclei red and, as only one colour is present, these areas can be selected on the basis of the colour intensity. The percentage area of cell cover is quickly calculated from this data. The data obtained is analysed using Minitab Statistical softwear (Minitab Inc).

BalbMK cells initially form small clusters on the surface of the collagen gel in a submerged culture (Figure 2). These clusters become less defined on prolonged culture as the cells spread out and form a more even monolayer (Figure 2). The morphology of the BalbMK cells in this model resembles a simplified epithelium. This model represents a normal "control" murine living skin equivalent.

SP-1 cells adhere to the surface of the collagen gel and spread out across the surface (Figure 3). After 5 days in culture the cells start to retract and by 8 days have formed distinct clusters (Figure 3). The cells stack to form clusters 3-4 cells thick. Raising the culture to the air interface promoted the formation of these "papilloma" structures. This model shows the benign papilloma stage of tumour progression.

T52 cells adhere to the surface of the collagen gel and spread out across the surface of a submerged culture (Figure 4). The cell cover decreases with time in culture (Figure 4). Viewing this model in cross section shows that the cells have invaded into the collagen gel after 3 days in culture (Figure 4). This model shows an invasive, malignant phenotype typical of a carcinoma.

Example 3: Reproducibility and viability of the models

Reproducibility studies were performed using both BalbMK and SP-1 models grown over a time course and harvested as submerged cultures at day 3 and day 4 and raised to the air interface at day 3, 4, 5 and 6. Models were set up in quadruplicate. Four fields of view were photographed for each of the 48 different models and the area of growth measured in each one. Mean areas and standard errors were calculated for each group. Fields of view compared from within the same gel gave similar areas of growth. Measurements made of different models, which were cultured under the same conditions, gave comparable results. There were instances where areas measured were different from their replicates. This was due primarily to the presence of a large cluster of cells. Replicate models are routinely set up for all treatments studied and several fields of view are studied

for each model to minimise errors introduced by natural variations and to act as a quality control.

Bias of the data and methods of trying to overcome this

As the cells on the surface of the models frequently form patterns or interesting morphologies there was a danger that a bias may be introduced when photographing the models to the areas of greatest interest. This has been overcome by photographing several fields of view at random followed by randomly selecting a subset of these in an attempt to minimise any bias introduced by the person analysing the data. The models often showed unusual cell distributions at the edge of the model. These areas were avoided when measurements were being made. Samples were recorded using number codes to minimise bias in the interpretation of the data.

Example 4: Use of the model to evaluate test agents

All-trans-Retinoic acid (RA) was selected as the first test agent for use in this system. RA is a well characterised agent which has been shown to inhibit mouse skin papilloma growth and has been used extensively in monolayer culture.

To test RA in the model, a single concentration of RA was selected and the models harvested over a time course. SP-1 models were set up and cultured in

submerged conditions for 4 days. The cultures were treated with 10^{-7}M RA and gels were harvested and fixed over a time course at days 6, 7, 10, 11 and 12, corresponding to 2, 3, 4, 5 and 6 days with RA respectively. 10^{-7}M is the concentration that is routinely used in monolayer cultures but as the three-dimensional *in vitro* models reflect many properties of *in vivo* tissue it is possible that higher concentrations may be required to achieve a similar effect.

A concentration gradient of RA was used in submerged cultures of the SP-1 model. The SP-1 cells were cultured on the collagen lattices and after four days of submerged culture RA was added to the cultures at 10^{-6}M, 10^{-7}M and 10^{-8}M with untreated SP-1 cells as the control. Models were harvested in replicate at 8 and 10 days of submerged culture (corresponding to 4 and 6 days with RA respectively) and the data was processed for analysis.

Morphological differences characteristic of apoptosis, such as size and change in nuclei composition were detected in all models using both the H & E and PI staining methods. This allows cell death by apoptosis to be quantified within the model system.

Growth curves of SP-1 cells in monolayer culture showed that retinoic acid had a growth inhibitory effect on SP-1 cells with 46% inhibition of the log phase of growth at 10 days in culture with 10^{-7}M RA. SP-1 cell

models treated with 10^{-7}M RA showed a modest effect with less cell cover than the corresponding untreated controls (Figure 5). Studying the effect of a concentration gradient of RA on the SP-1 model showed a marked effect on the model at the highest concentration studied, of 10^{-6}M RA, with a considerable decrease in cell cover (Figure 6). The degree of growth inhibition was shown to be concentration dependent. Incorporation of RA into the T52 Hufos model showed no effect (Figure 7). The T52 Hufos cells showed resistance to RA. These data demonstrate that the SP-1 and T52 Hufos models show different and independent behaviour to the test agent RA.

Example 5: Development of further models.

Three additional *in vitro* models have been constructed using mouse tumour cells to diversify the application of this model system for testing anti-cancer therapeutic agents.

All three cell lines tested were of epithelial origin. The CMT 93/69 mouse rectum carcinoma (Franks and Hemmings 1977), the CMT 64/61 mouse lung carcinoma (Franks *et al.* 1976) as described in Example 9 and TA3 Hauschka mouse mammary carcinoma cells (Hauschka 1953; Klein *et al.* 1972) were seeded onto collagen gels, which had been contracted with embryonic or newborn stromal cells, and cultured as submerged cultures for 6 days.

Figure 9, shows the visualisation of these models both as whole mounts and as H & E paraffin sections. CMT 93/69 rectal carcinoma cells produced an intact epithelial layer within this system (Figure 9B). A similar epithelial layer was also observed with the CMT 64/61 lung carcinoma cell line (Figure 9D). The mouse mammary carcinoma TA3 Hauschka produced an epithelium which was more clustered in appearance (Figures 9E and 9F) and some cellular invasion was observed.

Advantages of the system:

The present system provides an effective *in vitro* replacement for animal testing for new anti-tumour agents in a living mouse model and also provides a good replacement for current monolayer culture assays. The fibroblasts utilised in these models were obtained from the dermis of newborn mice, a single litter providing enough cells for circa 200 gels. The *in vitro* model has the advantage that it takes considerably less time than an animal model to yield data. The models are quick and easy to set up and require only days to produce the relevant papilloma and carcinoma models, in comparison with animal studies where several weeks are required before SP-1 cells generate a papilloma at the graft site. Shorter culture periods are desirable for productivity and rapid turnover of data.

Moreover, this novel method of analysis and detection allows effects to be observed in three dimensions, vertically and horizontally. This allows total cell coverage on the surface of the model to be quantified (horizontal two-dimensional measurements) as well as the amount of invasion into the support gel. Fluorescent vital cell labelling allows analysis (vertical three-dimension measurements) of cell dynamics in the living tumour models. This approach lends itself to simultaneous automated dynamic monitors of multiple tissue models for testing purposes.

Test agents can be introduced into the model once the benign (eg. SP-1 model) or carcinoma (eg. T52 Hufos model) structures have formed, and studied for their effect on established growth. Alternatively the test agent can be introduced at the earlier stages of development in the model. This flexibility of the model has the potential to provide valuable data on the mode of action of a specific test agent.

Used in combination, a benign model and malignant model can be used as a conversion assay to study the conversion of cells from the benign papilloma stage of development to the invasive, malignant stage. This has valuable implications for studying new therapeutic agents as it will aid in the determination as to what stage in tumour development a new drug requires to be

administered. Ultimately, this system will allow a wide range of new therapeutic agents to be evaluated for their efficiency as anti-cancer agents.

The model system has been designed to screen new compounds that have not been characterised previously with respect to their activity towards carcinogenic tissue. By testing the new agent in the presently described models, it could be provided to the customer with a breakdown of how this new agent will behave against benign "papilloma-like" cell clusters as well as against the latter malignant carcinoma stage. The diversity of this system will allow not only the changes in cell cover to be determined but also changes in the cell phenotypes, observations of cell death, differentiation, and invasion (or lack of it). The use of a combination of the benign and malignant models will allow the agent to be tested for its effectiveness during the conversion phase or a tumour from a benign to a malignant phenotype. This is of significant importance as some test agents may work more effectively at the early stages of tumour development rather than at the later stages. The implications of such a model for testing the potential therapeutic properties of a test agent is also valuable.

References:

Franks L.M., Carbonell A.W., Hemmings V.J., Riddle P.N. (1976). Metastasizing Tumours from Serum-supplemented and Serum-free Cell Lines from a C57BL Mouse Lung Tumor. Cancer Res. 36: 1049-1052

Franks L.M., Hemmings V.J. (1978). A cell line from an induced carcinoma of mouse rectum. J. Pathol. 124: 35-38.

Greenhalgh D.A., Yuspa S.H. (1988). Malignant Conversion of Murine Squamous Papilloma Cell Lines by Transfection with the *fos* Oncogene. Molecular Carcinogenesis 1: 134-143.

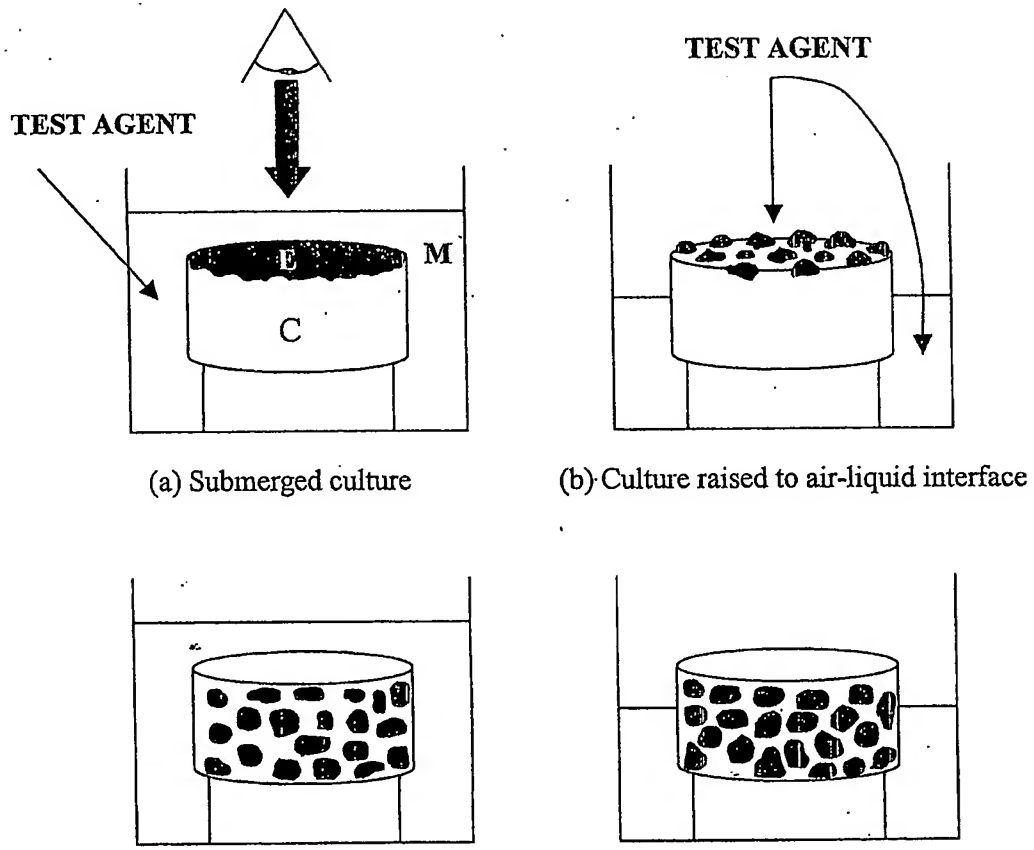
Hauschka T.S. (1953). Cell Population Studies on Mouse Ascites Tumors. Trans. N.Y. Acad. Sci. 16: 64

Klein G., Friberg S., Harris H. (1972). Two Kinds of Antigen Suppression in Tumor Cells Revealed by Cell Fusion. J. Exp. Med. 135: 839-849.

Strickland J.E., Greenhalgh D.A., Koceva-Chyla A., Hennings H., Restrepo C., Balaschak M., Yusap S.H. (1988). Development of Murine Epidermal Cell Lines Which Contain an Activated ras^{Ha} Oncogene and Form Papillomas in

Skin Grafts on Athymic Nude Mouse Hosts. Cancer Res 48: 165 - 169.

Weissman B., Aaronson S.A. (1985). Members of the *src* and *ras* Oncogene Families Supplant the Epidermal Growth Factor Requirement of BALB/MK-2 Keratinocytes and Induce Distinct Alterations in Their Terminal Differentiation Programme. Mol. Cell. Biol. 5: 3386-3396.



(a) Submerged culture

(b) Culture raised to air-liquid interface

(c) Submerged culture of gel containing both stromal and tumour cells

(d) Culture raised to air-liquid interface

Figure 1: The model (a) comprises a collagen gel (C) contracted by embryonic or newborn, organ specific stromal cells, which support a layer of either benign or malignant mouse epithelial cells (E). Models are submerged in media (M) for the initial stages of culture (a and c) and are thereafter raised to the air-liquid interface (b and d) to promote cell differentiation. Test agents with potential anti-cancer activity can be added into the model system at different stages of tumour development (a and b) either into the media of the submerged (a) and raised cultures (b) or onto the surface of the model in the raised culture (b). Models can also be prepared incorporating both stromal and tumour cells within the gel (c and d).

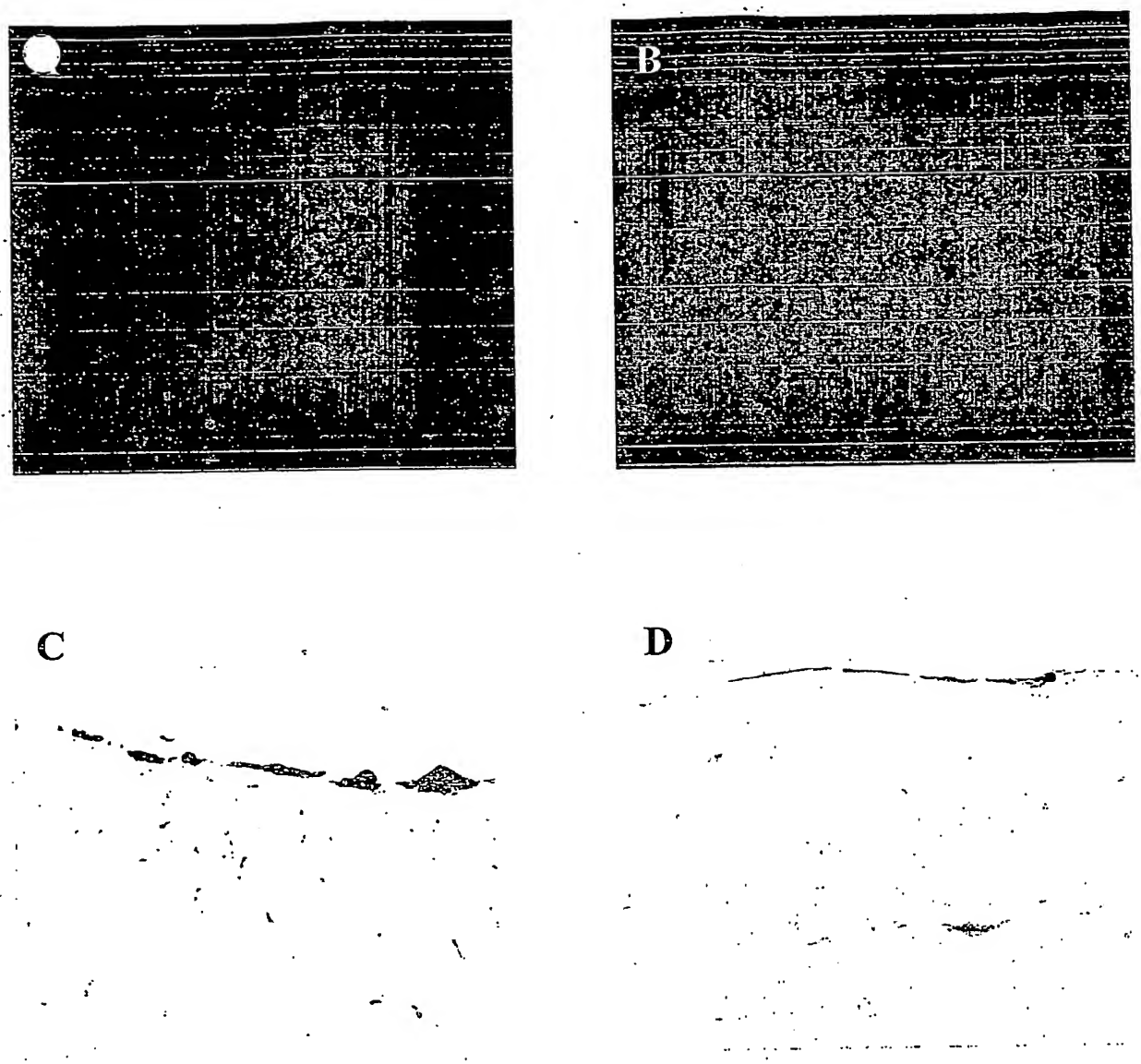


Figure 2: BalbMK "control" model:
Propidium iodide stained whole mounts (x 100 magnification) of (A) 2 day submerged culture and (B) 9 day submerged culture and H & E stained paraffin sections (x 100 magnification) of (C) 2 day submerged culture and (D) 10 day culture which has been raised to the air interface for 4 days.

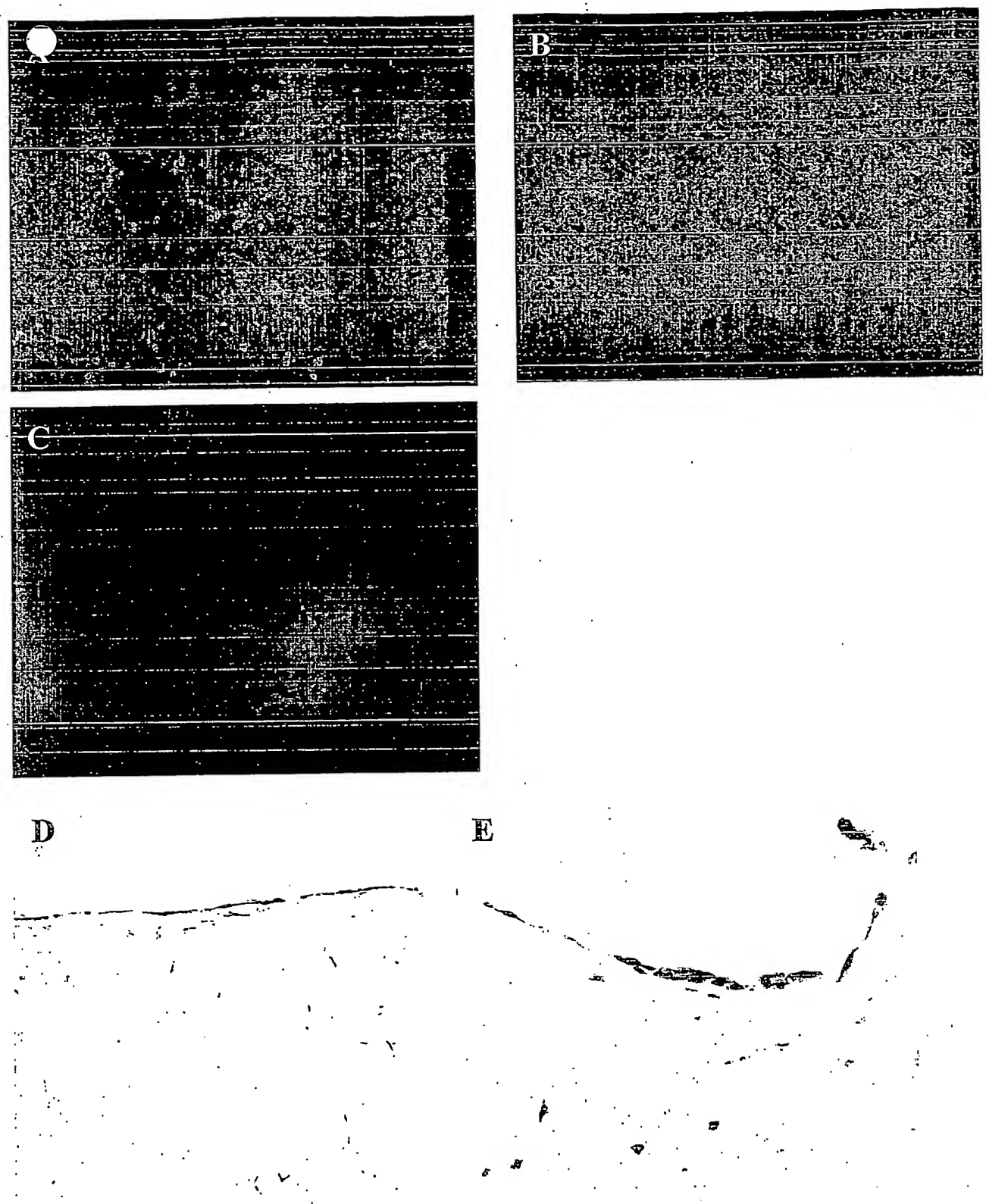


Figure 3:SP-1 "papilloma" model:
Propidium iodide stained whole mounts (x 100 maginification) of (A) 2 day submerged culture (B) 8 day submerged culture (C) 9 day culture which has been raised to the air interface for 1 day and H & E stained paraffin sections of (D) a 2 day submerged culture (x 100 magnification) and (E) a 10 day culture which has been raised to the air interface for 4 days.

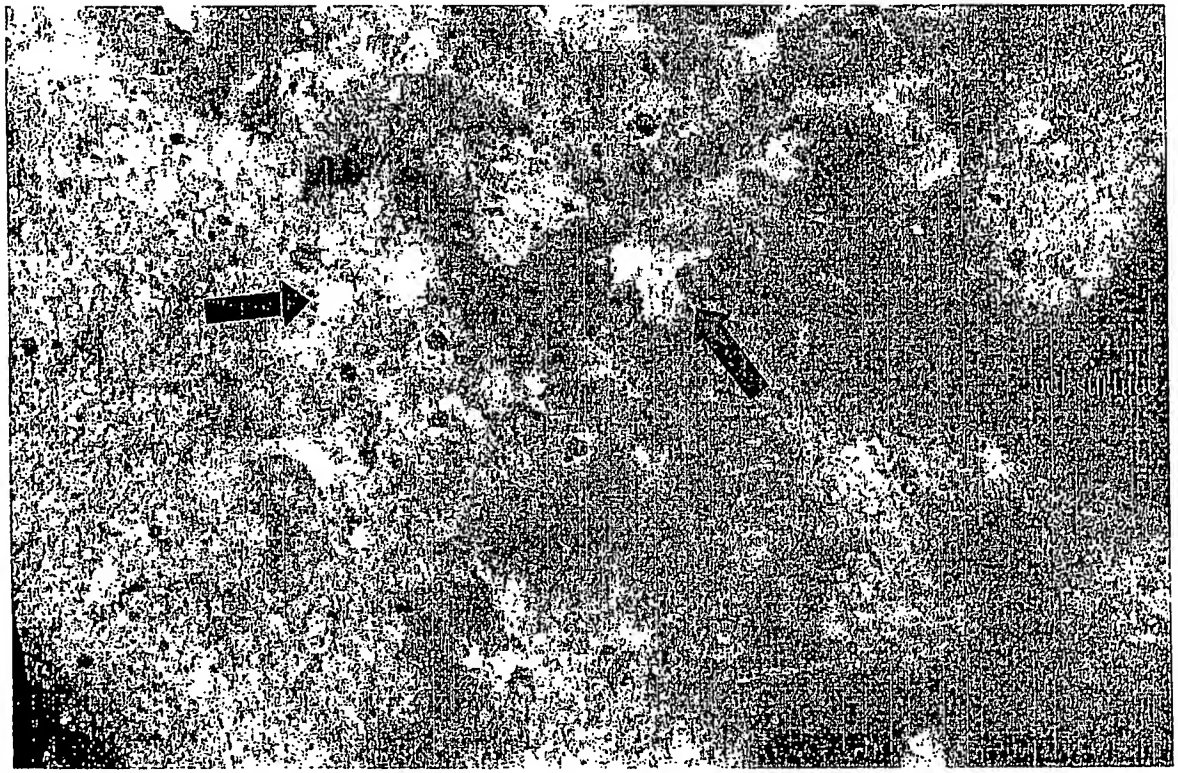


Figure 3 (F): Reconstructed three-dimensional confocal image of a model, showing the surface of the collagen gel, with SP-1 cells covering the surface of the gel and piling up into papilloma-like structures (black arrows).

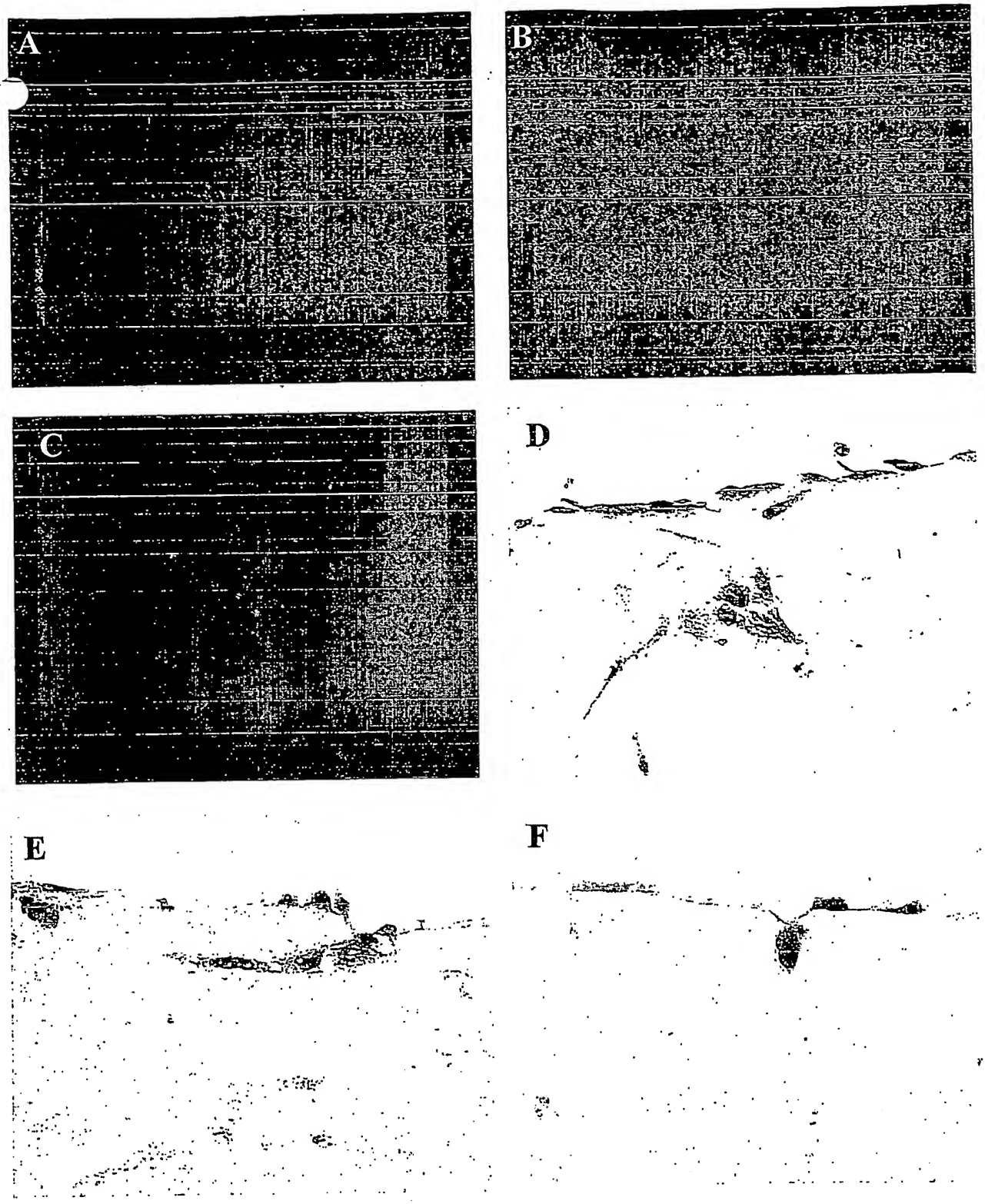
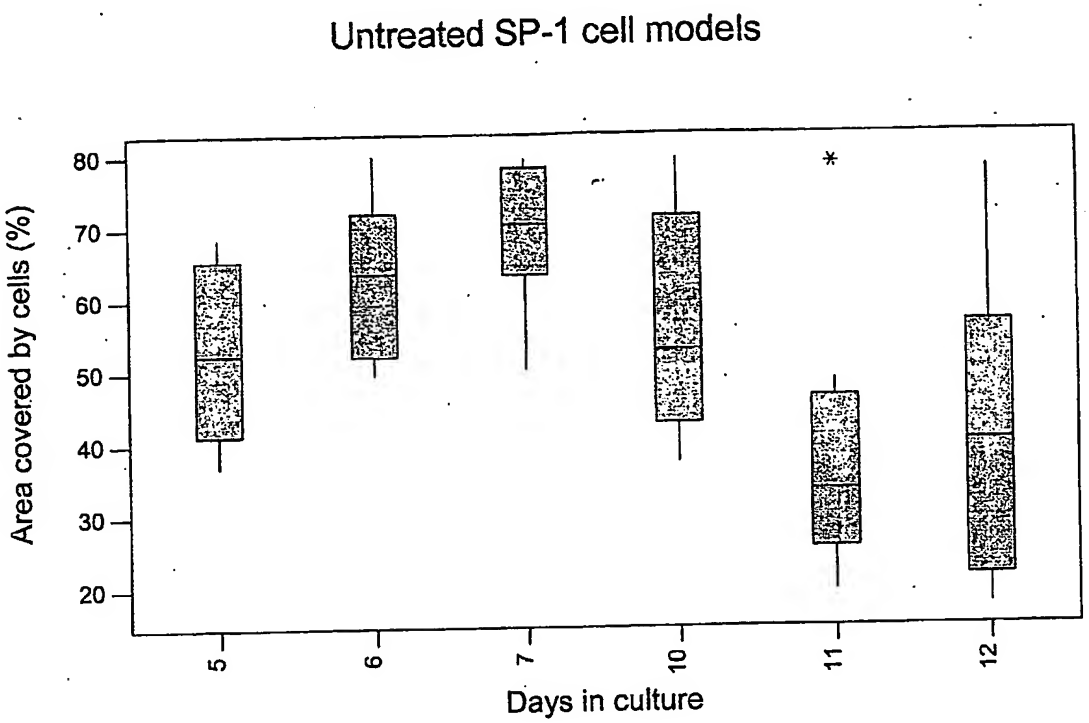


Figure 4: T52 Hufos "invasive tumour" model
Propidium iodide stained whole mounts (x 100 magnification) of (A) 4 day submerged culture (B) 6 day submerged culture (C) 10 day culture that has been raised to the air interface for 4 days and H & E paraffin stained sections (x 400 magnification) of (D) 4 day submerged culture (E) 6 day submerged culture and (F) 10 day culture which has been raised to the air interface for 4 days.

Figure 5: The effect of 10^{-7}M retinoic acid on the area of cell cover in the SP-1 papilloma model over a 12 day period.



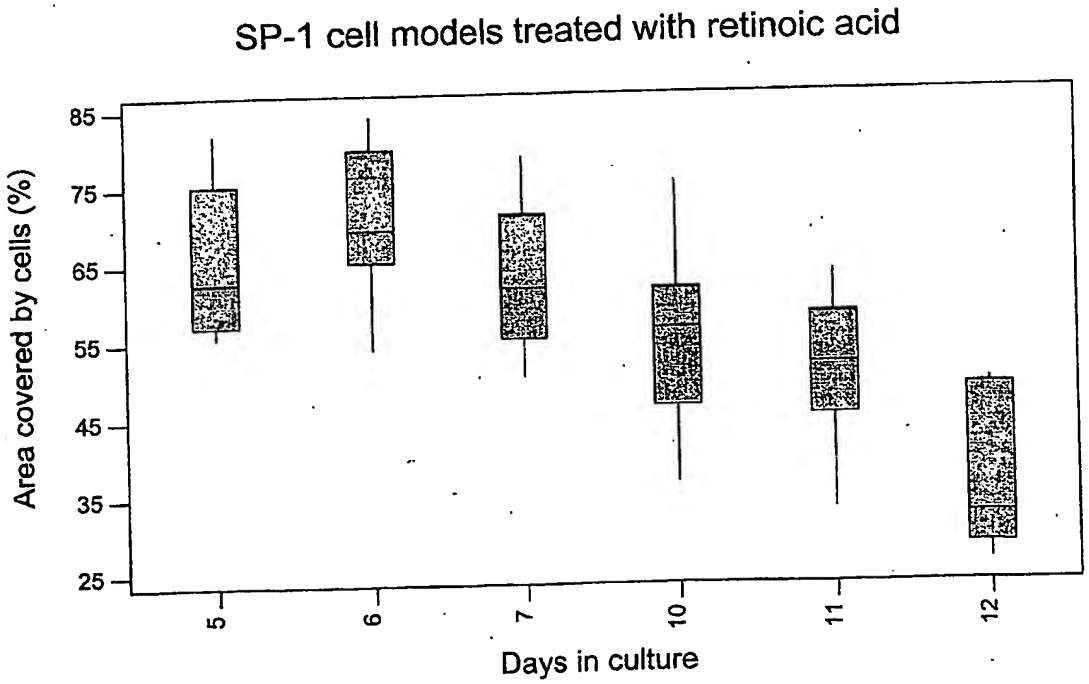
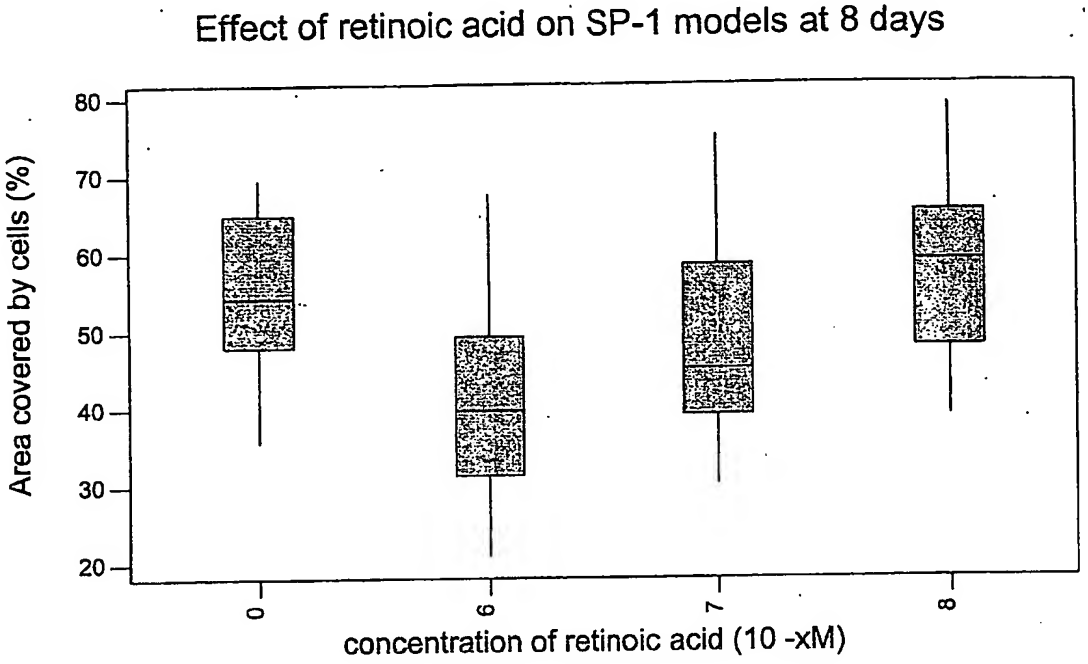


Figure 6: The effect of retinoic acid concentration on the area of cell cover in the SP-1 papilloma model at day 8 and day 10 of culture.



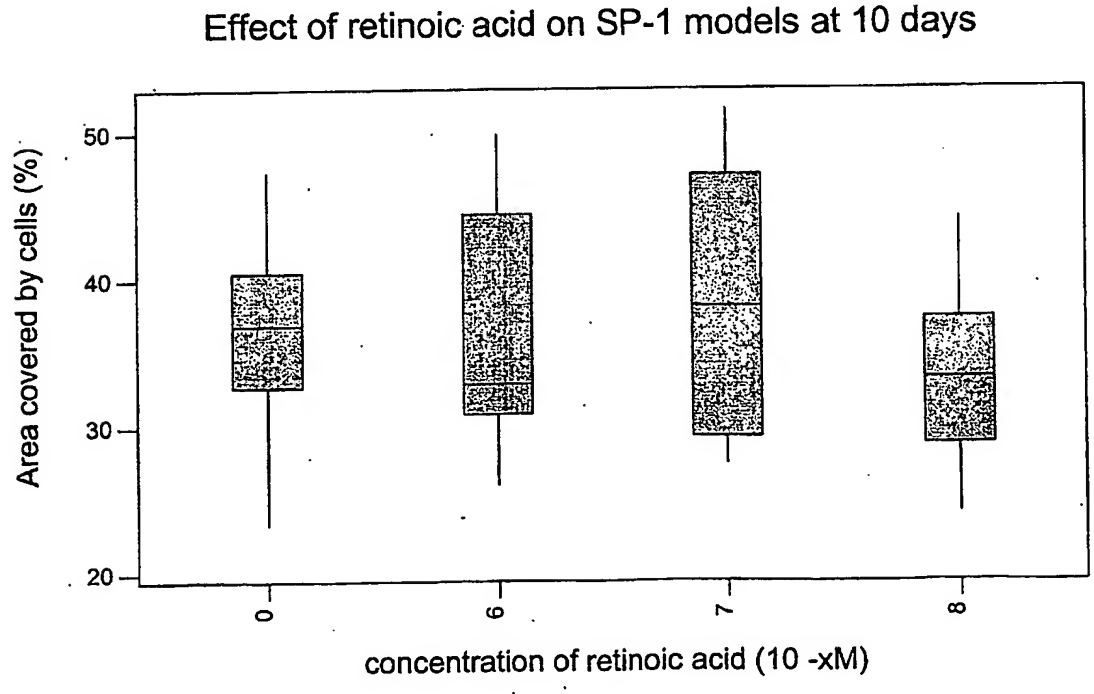
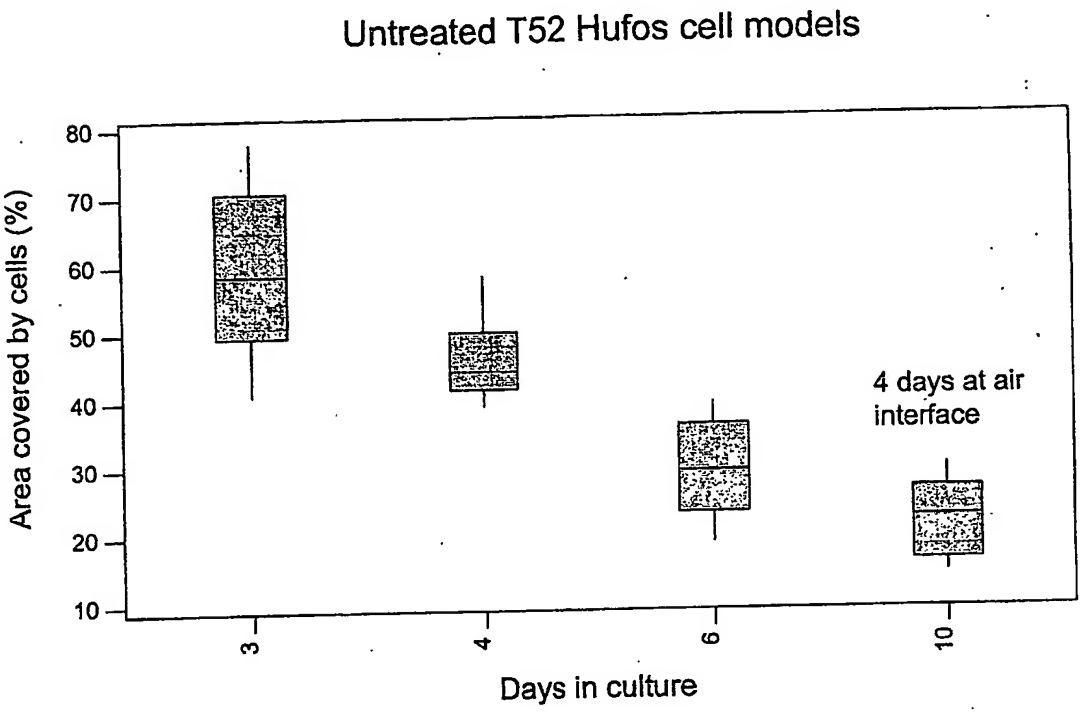
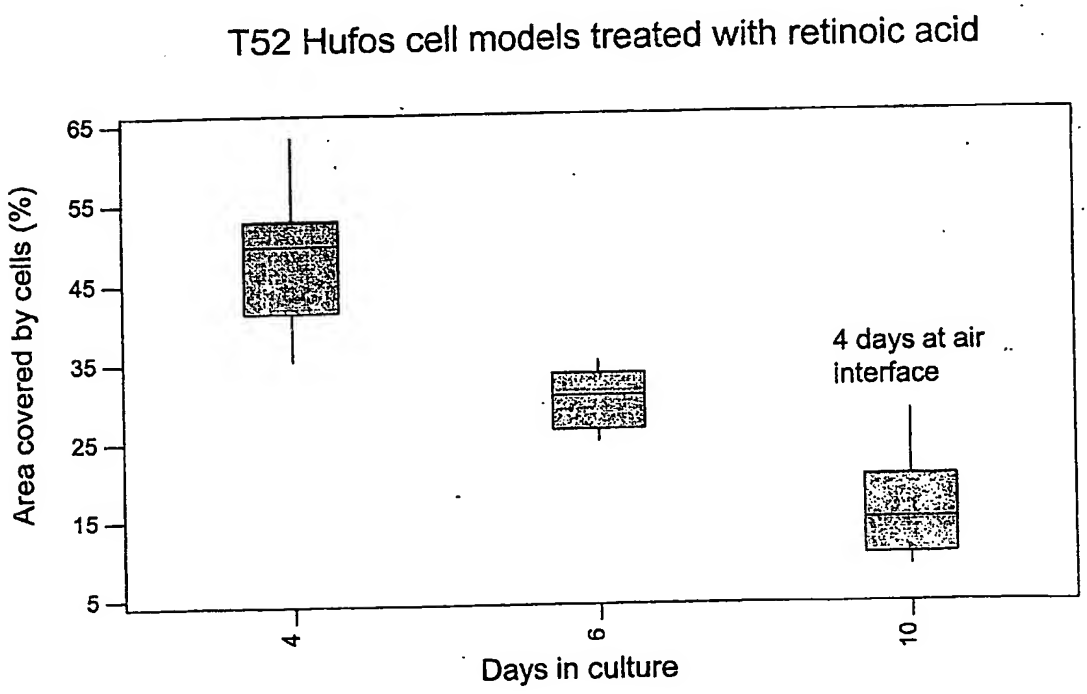


Figure 7: The effect of 10^{-7}M retinoic acid on the area of cell cover in the T52 Hufos invasive tumour model over a 10 day culture period.





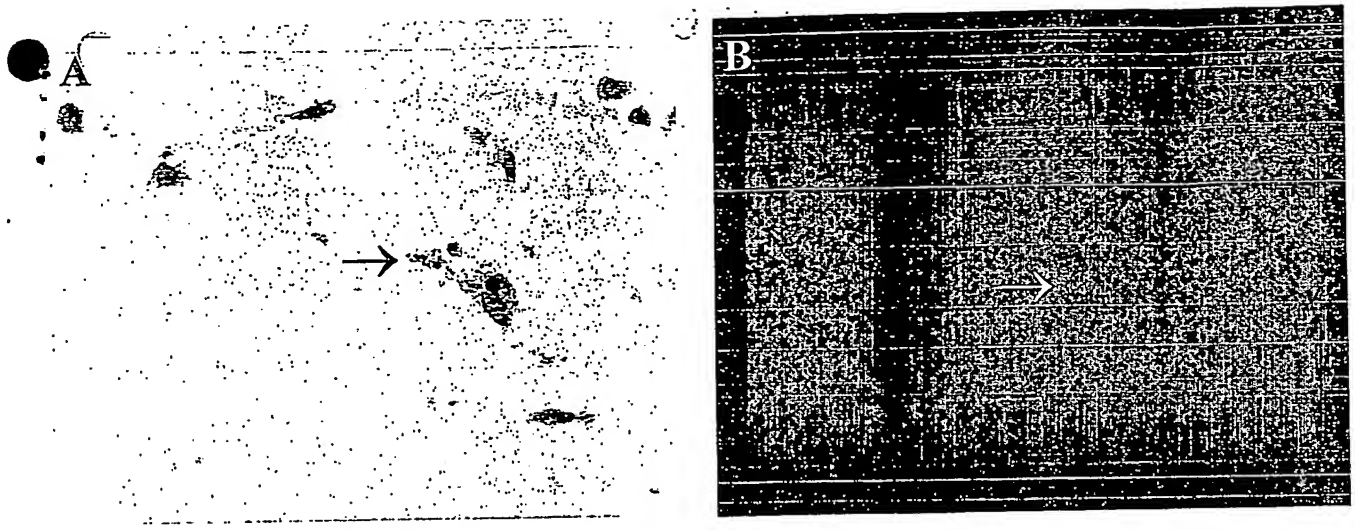


Figure 8: Detection of apoptotic cells.
Condensed and fragmented cell nuclei, characteristic of cell death by apoptosis, can be detected on (A) an H & E stained paraffin section of a T52 Hufos model (x 400 magnification) and (B) a propidium iodide whole mount of an SP-1 model (x 200 magnification).

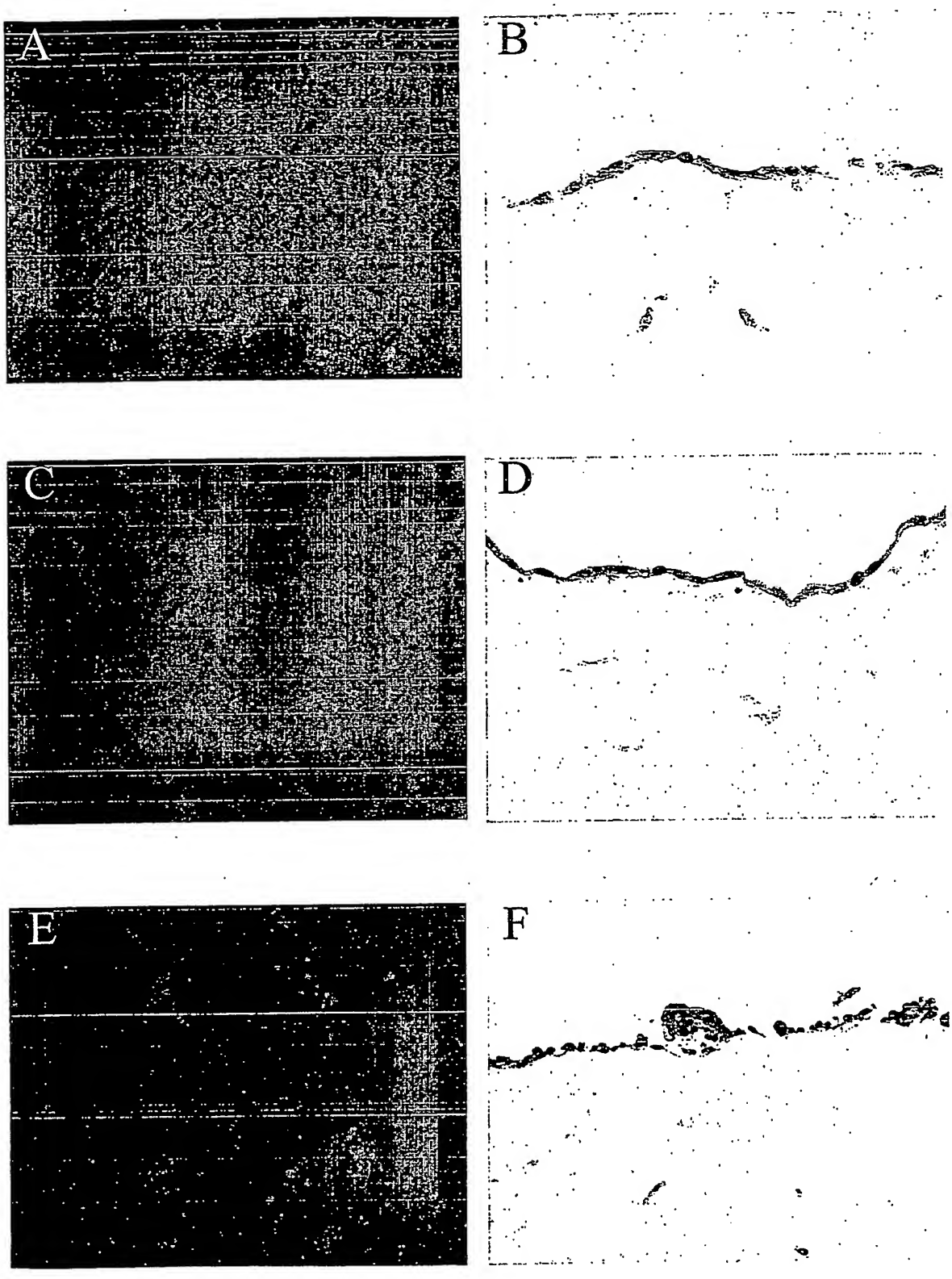


Figure 9: Vital dye stained whole mounts (x 100 mag) (A, C and E) and H &E stained paraffin sections (x 400 mag) (B, D and F) of submerged cultures of: CMT 93/69 mouse rectum carcinoma (A & B), CMT64/61 mouse lung carcinoma (C & D) and TA3 mouse mammary carcinoma (E and F).